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AMENDMENT TO THE SPECIFICATION

Please correct the published version of the specification as indicated below because the published version differs from that originally filed by the Applicants. The originally filed specification refers to "IFN α 2b" at page 37, line 17 but corresponding paragraph [0083] of US 2004/0223949 A1 incorrectly refers to "IFN γ ". Paragraph [0088] also contains a typographical error in that "Ionomycin" referred to at originally filed page 39, line 25 published as "Ionomycin". These corrections are being requested due to errors made solely by USPTO. Replacement paragraphs [0083] and [0088] are shown below:

[0083] Treatment with HDI: IFN γ α 2b (Schering Canada, Pointe-Claire, Quebec) was administered using the dose and schedule previously tested. (Kirkwood, et al. 1996. *J.Clin.Oncol.* 14, 7-17) HDI consisted of 20 MU/m²/d IV 5 days/week x 4 weeks. The IFN α 2b dose was held and then restarted at a 33% dose reduction if severe toxicity (grade 3 or 4, defined by the common toxicity criteria established by the National Cancer Institute Cancer Treatment Evaluation Program; Kirkwood, et al. 2001. *J.Clin.Oncol.* 19, 2370-2380) was observed. A second decrease of 33% of the original dosage was made in some patients for recurrent severe toxicity.

[0088] ELISPOT assays: HA-multiscreen plates (Millipore, Bedford, MA) were coated overnight at room temperature with 75 μ l of anti-IFN- γ mAb from the 1-DIK clone (MABTECH, Stockholm, Sweden) (2 μ g/ml in PBS). The plates were then washed with PBS, to remove unbound antibody, and blocked with 0.5% BSA/PBS for 1 h at room temperature. PBMC were added in duplicate or triplicate wells in the presence or absence of peptide. The two modified gp100 peptides were added at a final concentration of 25 μ g/ml and the FLU peptides were added at a final concentration of 10 μ g/ml. Mitogenic stimulation was performed with phorbol myristic acetate (PMA) (20 ng/ml) (Sigma) and Ionomycin (1 μ g/ml) (Calbiochem, San Diego, CA). IL-2 (100 IU/ml) was included in all cultures unless stimulated by mitogens. After incubation at 37°C in 5% CO₂ for 24 h, the cells were discarded, and the plates were washed extensively with 0.05% Tween/PBS. Secondary biotinylated anti-IFN- γ mAbs (clone 7-B6-1,

MABTECH) were then added (75 μ l/well at 1 μ g/ml) and left for 2 h at room temperature, followed by extraavidin-conjugated alkaline phosphatase (Sigma) for an additional 1 h. The plates were developed using NBT/BCIP phosphatase substrate solution (Sigma) and counted using a stereomicroscope at 40x and an automated ELISPOT reader (Carl Zeiss Vision, Germany). Statistical analysis was carried out using Microsoft Excel software.